National Institute for Health and Care Excellence

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CVD risk assessment and lipid modification

Network meta-analysis for change in LDL-C and non-HDL-C

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1 Introduction

2 Network meta-analysis (NMA) is a statistical technique that allows simultaneous pooling of 3 data for three or more interventions when the available evidence forms a connected network 4 of intervention comparisons from RCTs. This enables both direct evidence and indirect 5 evidence to be pooled. NMA combines all the available data simultaneously into a single set 6 of treatment effects that provide a unique ordering of intervention effectiveness, whilst 7 respecting the randomisation in the included RCTs. The resulting estimates are therefore 8 easier to interpret than a series of pairwise comparisons, enables ranking of the 9 interventions, and because both direct and indirect evidence is pooled treatment effects are 10 more precisely estimated (have greater statistical power).

11 The analysis provides estimates of relative effects (with 95% credible intervals) for each 12 intervention compared to a reference intervention (in this case the reference intervention was 13 placebo, which included background statin treatment) as well as estimates of all pairwise 14 comparisons. In addition, for a given assumed "baseline effect" on the reference intervention, 15 we can obtain absolute effects for all interventions. These estimates provide a useful clinical 16 summary of the results and facilitate the formation of recommendations based on the best 17 available evidence. Having a single set of intervention effects that takes into account all the 18 available evidence also facilitates cost effectiveness analysis.

19 The review for this guideline update (comparing adding ezetimibe, PCSK9 inhibitors or 20 inclisiran to statins with statins alone) formed a connected network of RCT evidence and so 21 an NMA was considered. This topic was considered a high clinical priority due to variations in 22 practice and uncertainty about the most clinically and cost-effective strategy. It was also 23 given the highest priority for new economic modelling as direct evidence on the relative 24 effectiveness of different treatment options could not inform the optimal lipid level treatment 25 target. Given this, the committee agreed that network meta-analysis was warranted to 26 facilitate cost effectiveness analysis and help decision making in this area.

2 Study selection

2 A systematic review of RCTs comparing licenced doses of ezetimibe, inclisiran, alirocumab

3 or evolocumab with each other, high or medium intensity stains, usual care or placebo in

4 adults with CVD was undertaken for the guideline. Studies identified in this review were

5 considered for inclusion in the NMA.

6 We performed NMAs that simultaneously used all the relevant RCT evidence from the

7 clinical evidence review. As with conventional meta-analyses, this type of analysis does not 8 break the randomisation of the evidence.

9 Further details of the evidence identified from the review can be found in the evidence review 10 and the protocol, evidence tables and GRADE assessment appendices.

11 2.1 NMA model assumptions

12 2.1.1 Class effect models of evolocumab and alirocumab

13 Three different approaches to modelling dose and class effects for alirocumab and evolocumab were explored. Goodness of fit was assessed using the posterior mean of the residual deviance, where a well-fitting model would show total residual deviance equivalent to the number of data points. Different models of the class effect structure were compared on the basis of DIC, which is a measure of fit penalised for model complexity, residual deviance and estimates of heterogeneity (between-study and within-class standard deviation).

19 The simplest model to assume that all treatments and doses have the same relative effect 20 compared with placebo; that there is a common class effect (sometimes called a fixed class 21 effect model). Any differences would be captured as between study variability. The second 22 model assumed that treatment and dose effects differ, so a mean effect across treatments 23 and doses within class was estimated, with between treatment/dose variability (a random 24 class effect model). The third model assumed a common (fixed) class effect for the two 25 treatments but allowed for an effect of high and low dose for evolocumab. Of these three 26 models, the common class effect (fixed class effect) model gave the best fit for each 27 outcome and dataset, and so all results are reported using this model.

28 2.1.2 Network meta-regression on baseline lipid level

Network meta-regression was conducted using 2 different models to explore whether differences in mean lipid levels at baseline interacted with the relative treatment effects. The simplest was to assume that mean lipid levels moderated the effect of each active treatment in the same way relative to non-active treatments. This assumption was then relaxed to estimate a different covariate effect for each treatment class. There was very limited evidence with which to estimate the meta-regression models, and the effects are very uncertain, but there was no evidence of effect modification by baseline mean lipid levels in the studies. Specifically, the large credible intervals around the meta-regression term for baseline severity, and the fact that they include zero mean that it is not possible to infer an effect of baseline lipid level for these data. Therefore, in agreement with the committee, the results from this analysis were not used as the base case.

40 **2.2 Outcome measures**

41 Network models were fitted for the following outcomes:

- Percentage change from baseline in LDL cholesterol (%)
- 43 Absolute change from baseline in LDL cholesterol (mmol/litre)

- Percentage change from baseline in non-HDL cholesterol (%)
- Absolute change from baseline in non-HDL cholesterol (mmol/litre)

3 Studies varied in whether they reported the outcomes as a percentage change from baseline

4 or as an absolute outcome change from baseline, and some studies reported both. It is not

5 possible to combine these two different outcome measure formats, and so we conducted

6 NMA for both outcome formats separately.

3 Results

2 3.1 Imputed correlations and standard deviations

3 The correlation between lipid levels at baseline and follow-up was estimated to be 0.386 from

4 14 studies reporting mean LDL at baseline, follow-up and the change from baseline. This

5 was used to estimate the standard error for mean change from baseline for studies that

6 report baseline and follow-up means only. Imputed SD was specific to lipid type and outcome

7 format (**Table 1**) and were used for studies which did not report a standard deviation or

8 standard error.

9 Table 1: Values of imputed SD by data type

Dataset	Mean SD (imputed)
LDL, reported as percent change	24.87
LDL, reported in units of mmol/L	0.751
Non-HDL, reported as percent change	23.01
Non-HDL, reported in units of mmol/L	0.9574

10 3.2 Percentage change from baseline in LDL cholesterol

11 3.2.1 Network and data

12 Eighteen studies reported in 17 papers^{1-4, 6, 7, 9-12, 16, 17, 21-23, 26, 28} were identified as reporting 13 outcome data for percentage change from baseline in LDL cholesterol. Eight treatments were 14 included in the network: placebo (including placebo and/or statin treatment), standard care, 15 ezetimibe, inclisiran, alirocumab, evolocumab (mixed), evolocumab 140mg and evolocumab 16 420mg. Alirocumab and evolocumab were modelled as a single fixed class. The network can 17 be seen in **Figure 1** and the trial data for each of the studies included in the NMA are 18 presented in **Table 2**.

1 Figure 1: Network diagram for percentage change in LDL cholesterol



3

4 Table 2: Study data for percentage change in LDL cholesterol at 3-12 months network 5 meta-analysis

			% change LDL-C	
Study	Intervention	Comparison	Mean differe nce	SE
Cannon, 2015 ³ IMPROVE-IT	Ezetimibe	Placebo	-24	0.423
Hougaard, 2017 ⁹	Ezetimibe	Placebo	-9.6	3.513
Joshi, 2017 ¹⁰	Ezetimibe	Placebo	-11.9	3.932
Masuda 2015 ¹⁶	Ezetimibe	Placebo	-19	5.984
Ran, 2017 ²¹ NSTE-ACS	Ezetimibe	Placebo	-23.4	5.427
Tsujita, 2015 ²⁸ PRECISE-IVUS	Ezetimibe	Placebo	-11	2.981
Kereiakes, 2015 ¹¹ ODYSSEY COMBO I	Alirocumab	Placebo	-41	3.678
Koh, 2018 ¹² ODYSSEY KT	Alirocumab	Placebo	-63.4	4.173
McCullough, 2018 ¹⁷	Alirocumab	Placebo	-61.8	1.442

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			% change LDL-C	
Study	Intervention	Comparison	Mean differe nce	SE
ODYSSEY-LONG TERM				
Ray, 2019 ²² DM-INSULIN	Alirocumab	Placebo	-48.5	4.4
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Evolocumab low	Placebo	-66.1	2.759
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Evolocumab high	Placebo	-50.3	2.886
Sabatine, 2017 ²⁶ FOURIER	Evolocumab	Placebo	-59	0.510
Ray, 2020 ²³ ORION-10	Inclisiran	Placebo	-49.2	1.224
Ray, 2020 ²³ ORION-11	Inclisiran	Placebo	-53.8	1.249
Ako, 2019 ¹ ODYSSEY J-IVUS	Alirocumab	Standard care	-50.5	2.758623
Ray, 2019 ²² DM-DYSLIPEDIMIA	Alirocumab	Standard care	-45.9	5.8
Cannon, 2015 ⁴ ODYSSEY COMBO II	Alirocumab	Ezetimibe	-31.2	2.581
Han, 2020 ⁷ ODYSSEY EAST	Alirocumab	Ezetimibe	-35.7	2.535

1 3.2.2 Results of network meta-analysis

2 **Table 3** summarises the results of the pairwise meta-analyses in terms of mean differences

3 generated from studies directly comparing different interventions, together with the results of

4 the NMA (this is from a random effects model, which was chosen on the basis of model fit), 5 in terms of mean differences comparing each available treatment option with placebo/statin.

6 Table 3: Risk differences for percentage change in LDL cholesterol at 3-12 months; 7 direct pairwise meta-analysis results and NMA results

			NMA random effects model – mean difference (95% credible intervals)		
Intervention	Comparison	Pairwise estimates - mean difference (95% confidence intervals)	Base case model	Model with meta- regression on baseline lipid levels	
Standard care	Placebo/ statin	NA	-6.34 (-19.40, 6.98)	-5.27 (-19.63, 9.49)	
Ezetimibe		-11.5 (-15.66, -7.33) ^a	-17.83 (-23.74, -11.89)	-26.29 (-57.75, 7.20)	
Inclisiran		-51.45 (-53.17, -49.74) ^b	-51.27 (-61.88, -40.52)	-59.01 (-87.26, -28.88)	
PCSK9i		-54.62 (-59.28, -49.97) ^b	-55.01 (-60.33, -49.39)	-61.37 (-90.66, -30.03)	
Inclisiran PCSK9i	ata madal	-51.45 (-53.17, -49.74) ^b -54.62 (-59.28, -49.97) ^b	-51.27 (-61.88, -40.52) -55.01 (-60.33, -49.39)	-59.01 (-87.26, -28.88) -61.37 (-90.66, -30.03)	

8 (a) Fixed effects model 9 (b) Random effects model

1 3.2.3 Inconsistency and goodness of fit

2 Both fixed effects and random effects baseline models were fitted to the data from the

3 studies. Moderate heterogeneity was observed for this outcome, with a large reduction in

4 residual deviance and DIC seen in the RE NMA model, relative to the FE NMA model (Table

5 4). This, together with the moderate between-study SD supports choice of the RE model

6 structure.

7 An inconsistency model was run and the model fit statistics were as seen in **Table 4**. The

8 NMA model has a slightly smaller DIC suggesting that there is no evidence of inconsistency.

9 Table 4: Model fit statistics – percentage change in LDL cholesterol at 3-12 months

Relative effect models ^(a)	Between study SD (95% Crl)	Meta-regression covariate on baseline lipid level	Deviance information criterion (DIC)	Total residual deviance
NMA Fixed effects	-	-	181.9	108.2
NMA (Random effects)	7.23 (4.49, 11.40)	-	105.2	19.0
Unrelated mean- effects (Random effects)	7.30 (4.42, 11.79)	-	105.4	19.1
Meta-regression (Random effects)	7.55 (4.31, 12.75)	2.82 (-7.40, 12.40)	98.5	17.0

(a) Number of data points in the NMA and inconsistency models (n=19). Number of data points in the metaregression model models (n=17).

10 The global check for inconsistency, modelling treatment effects independently of each other,

11 indicates that there is little inconsistency in this dataset, with similar model fit in the RE NMA 12 and RE UME models, and no evidence of local inconsistency. Figure 2 presents a dev-dev 13 plot, which shows the contributions of each study datapoint to the residual deviance under 14 the random effects UME and NMA models. There is no evidence of inconsistency, as there 15 are no points notably below the line of equality, which would be indicative of data better 16 predicted by the inconsistency model.

17 Taken together, the model fit and dev-dev plots suggest there was little evidence of

18 inconsistency in these data.

19 Figure 2: Contribution of each study datapoint to the residual deviance under the

20 random effects UME and NMA models – percentage change in LDL cholesterol at 3-12 21 months



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1 3.2.4 Sensitivity analyses and subgroup analyses

Within the sensitivity analysis, including studies reporting baseline LDL, adding a metaregression term on baseline LDL levels resulted in a model with good model fit (total residual
deviance was 17, relative to 17 data points). However, the effect of baseline LDL was
uncertain, 2.82 (95% credible interval (CrI): -7.40, 12.40), with the interval including zero.
Between-study SD was higher in the meta-regression model, indicating greater differences
between studies estimating the same treatment effect.

8 3.3 Absolute change from baseline in LDL cholesterol

9 3.3.1 Network and data

10 Thirty studies^{1-6, 8-16, 18-21, 23-33} were identified as reporting outcome data for absolute change

11 from baseline in LDL cholesterol. Eight treatments were included in the network: placebo

12 (including placebo and/or statin treatment), standard care, ezetimibe, inclisiran, alirocumab,

13 evolocumab (mixed), evolocumab 140mg and evolocumab 420mg. Alirocumab and

14 evolocumab were modelled as a single fixed class. The network can be seen in Figure 3 and

15 the trial data for each of the studies included in the NMA are presented in Table 5.

16 Figure 3: Network diagram for absolute change in LDL cholesterol



1 Table 5: Study data for absolute change in LDL cholesterol (mmol/litre) at 3-12 2 months network meta-analysis

			Absolute change LDL-C (mmol/L)	
			Mean	
Study	Intervention	Comparison	nce	SE
Arimura, 2012 ²	Ezetimibe	Placebo	-0.34	0.129
Cannon, 2015 ³ IMPROVE-IT	Ezetimibe	Placebo	-0.43	0.034
Hougaard, 2017 ⁹	Ezetimibe	Placebo	-0.10	0.181
Joshi, 2017 ¹⁰	Ezetimibe	Placebo	-0.57	0.129
Kouvelos, 2013 ¹³	Ezetimibe	Placebo	-0.43	0.168
Luo 2014 ¹⁴	Ezetimibe	Placebo	-0.40	0.121
Luo 2016 ¹⁵	Ezetimibe	Placebo	-0.56	0.093
Masuda 2015 ¹⁶	Ezetimibe	Placebo	-0.69	0.217
Ran, 2017 ²¹ NSTE-ACS	Ezetimibe	Placebo	-0.80	0.160
Ren, 2017 ²⁵	Ezetimibe	Placebo	-0.37	0.183
Tsujita, 2015 ²⁸ PRECISE-IVUS	Ezetimibe	Placebo	-0.30	0.092
Ueda, 2017 ²⁹ ZIPANGU	Ezetimibe	Placebo	-0.36	0.128
Wang 2016 ³¹	Ezetimibe	Placebo	-0.62	0.238
Wang 2017 ³⁰	Ezetimibe	Placebo	-0.45	0.154
West 2011/2011a ^{32, 33}	Ezetimibe	Placebo	-0.62	0.280
Kereiakes, 2015 ¹¹ ODYSSEY COMBO I	Alirocumab	Placebo	-1.10	0.093
Koh, 2018 ¹² ODYSSEY KT	Alirocumab	Placebo	-1.56	0.112
Raber, 2022 ²⁰ PACMAN-AMI	Alirocumab	Placebo	-1.41	0.116
Schwartz, 2018 ²⁷ ODYSSEY OUTCOMES	Alirocumab	Placebo	-1.24	0.013
Sabatine, 2017 ²⁶ FOURIER	Evolocumab	Placebo	-1.45	0.013
Giugliano, 2012 ⁶ LAPLACE-TIMI 57a	Evolocumab 140mg	Placebo	-2.04	0.100
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Evolocumab 420 mg	Placebo	-1.58	0.100
Nicholls, 2016 ¹⁹ GLAGOV	Evolocumab 420 mg	Placebo	-1.46	0.059
Nicholls, 2022 ¹⁸ HUYGENS	Evolocumab 420 mg	Placebo	-1.52	0.181
Ray, 2020 ²³ ORION-10	Inclisiran	Placebo	-1.38	0.033
Ray, 2020 ²³ ORION-11	Inclisiran	Placebo	-1.26	0.032
Ako, 2019 ¹	Alirocumab	Standard care	-1.24	0.071

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			Absolute change LDL-C (mmol/L)	
Study	Intervention	Comparison	Mean differe nce	SE
ODYSSEY J-IVUS				
Gao, 2021 ⁵	Alirocumab	Standard care	-0.76	0.141
Rehberger, 2022 ²⁴	Alirocumab, evolocumab	Standard care	-1.50	0.206
Cannon, 2015 ⁴ ODYSSEY COMBO II	Alirocumab	Ezetimibe	-0.90	0.073
Hao, 2022 ⁸	Evolocumab and ezetimibe	Ezetimibe	-0.71	0.095

1 3.3.2 Results of network meta-analysis

2 Table 6 summarises the results of the pairwise meta-analyses in terms of mean differences

- 3 generated from studies directly comparing different interventions, together with the results of
- 4 the NMA (this is from a random effects model, which was chosen on the basis of model fit),
- 5 in terms of mean differences comparing each available treatment option with placebo/statin.

6 Table 6: Risk differences for absolute change in LDL cholesterol at 3-12 months; 7 direct pairwise meta-analysis results and NMA results

			NMA random effects model – mean difference (95% credible intervals)	
Intervention	Comparison	Pairwise estimates - mean difference (95% confidence intervals)	Base case model	Model with meta- regression on baseline lipid levels
Standard care	Placebo/ statin	NA	-0.30 (-0.59, -0.01)	-0.28 (-0.60, 0.03)
Ezetimibe		-0.41 (-0.47, -0.34) ^a	-0.46 (-0.58, -0.34)	-0.30 (-0.95, 0.34)
Inclisiran		-1.32 (-1.37, -1.28) ^a	-1.32 (-1.60, -1.05)	-1.17 (-1.78, -0.58)
Evolocumab plus ezetimibe		NA	-1.17 (-1.62, -0.74)	-1.01 (-1.80, -0.23)
PCSK9i		-1.51 (-1.65, -1.37) ^b	-1.46 (-1.60, -1.33)	-1.30 (-1.91, -0.72)

8 (a) Fixed effects model

9 (b) Random effects model

10 3.3.3 Inconsistency and goodness of fit

Both fixed effects and random effects baseline models were fitted to the data from the
studies. Large heterogeneity was observed for this outcome, with a large reduction in
residual deviance and DIC seen in the RE NMA model, relative to the FE NMA model. This,
together with the moderate between-study SD (on the scale of the outcome) supports the

15 choice of the RE model structure.

16 An inconsistency model was run and the model fit statistics were as seen in Table 7.

- 17
- 18
- 19

1 Table 7: Model fit statistics – absolute change in LDL cholesterol at 3-12 months

Relative effect models ^(a)	Between study SD (95% Crl)	Meta-regression covariate on baseline lipid level	Deviance information criterion (DIC)	Total residual deviance
NMA Fixed effects	-	-	156.2	239.9
NMA (Random effects)	0.19 (0.12, 0.27)	-	-33.4	31.7
Unrelated mean- effects (Random effects)	0.19 (0.12, 0.28)	-	-32.4	32.1
Meta-regression (Random effects)	0.20 (0.13, 0.30)	-0.06 (-0.25, 0.14)	-26.9	29.8

(a) Number of data points in the NMA and inconsistency models (n=31). Number of data points in the metaregression model models (n=29).

2 The global check for inconsistency indicates that there is little inconsistency in this dataset,

3 with similar model fit in the RE NMA and RE UME models, and no evidence of local

4 inconsistency. Figure 4 presents a dev-dev plot, which shows the contributions of each study

5 datapoint to the residual deviance under the random effects UME and NMA models. There is

6 no evidence of inconsistency, as there are no points notably below the line of equality, which

7 would be indicative of data better predicted by the inconsistency model.

8 Taken together, the model fit and dev-dev plots suggest there was little evidence of 9 inconsistency in these data.

10 Figure 4: Contribution of each study datapoint to the residual deviance under the

11 random effects UME and NMA models – absolute change in LDL cholesterol at 3-12 12 months



14 3.3.4 Sensitivity analyses and subgroup analyses

15 Within the sensitivity analysis, including studies reporting baseline LDL, adding a meta-

16 regression term on baseline LDL levels gave a model with good statistical fit, but the effect of

17 baseline LDL was uncertain, -0.06 (95% Crl: -0.25, 0.14), with the interval including zero.

3.4 Percentage change from baseline in non-HDL cholesterol

3 3.4.1 Network and data

4 Thirteen studies reported in 11 papers^{1, 4, 6, 7{McCullough, 2018 #215, 11, 12, 16, 22, 23, 26} were identified as

- 5 reporting outcome data for percentage change from baseline in non-HDL cholesterol. Eight
- 6 treatments were included in the network: placebo (including placebo and/or statin treatment),
- 7 standard care, ezetimibe, inclisiran, alirocumab, evolocumab (mixed), evolocumab 140mg
- 8 and evolocumab 420mg. Alirocumab and evolocumab were modelled as a single fixed class.

9 The network can be seen in Figure 5 and the trial data for each of the studies included in the

10 NMA are presented in Table 8 and Table 2.

11 Figure 5: Network diagram for percentage change in non-HDL cholesterol



13

14 Table 8:Study data for percentage change in non-HDL cholesterol at 3-12 months15network meta-analysis

			% change LDL-C	
Study	Intervention	Comparison	Mean differe nce	SE
Masuda 2015 ¹⁶	Ezetimibe	Placebo	-15.5	5.67
Kereiakes, 2015 ¹¹	Alirocumab	Placebo	-29.1	3.34

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				% change LDL-C		
			Mean			
Study	Intervention	Comparison	nce	SE		
ODYSSEY COMBO I						
Koh, 2018 ¹² ODYSSEY KT	Alirocumab	Placebo	-51.5	3.46		
McCullough, 2018 ¹⁷ ODYSSEY-LONG TERM	Alirocumab	Placebo	-52.1	1.22		
Ray, 2019 ²² DM-INSULIN	Alirocumab	Placebo	-37.4	3.90		
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	iano, 2012 ⁶ Evolocumab low Placebo ACE-TIMI 57		-61.4	2.50		
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Evolocumab high	Placebo	-47.6	2.60		
Sabatine, 2017 ²⁶ FOURIER	Evolocumab	Placebo	-51.6	0.28		
Ray, 2020 ²³ ORION-11	Inclisiran	Placebo	-47.3	1.16		
Ray, 2020 ²³ ORION-10	Inclisiran	Placebo	-43.4	1.14		
Ako, 2019 ¹ ODYSSEY J-IVUS	Alirocumab	Standard care	-40.4	2.40		
Ray, 2019 ²² DM-DYSLIPEDIMIA	Alirocumab	Standard care	-31.1	4.30		
Cannon, 2015 ⁴ ODYSSEY COMBO II	Alirocumab	Ezetimibe	-22.9	2.08		
Han, 2020 ⁷ ODYSSEY EAST	Alirocumab	Ezetimibe	-27.6	2.08		

1 3.4.2 Results of network meta-analysis

2 Table 9 summarises the results of the pairwise meta-analyses in terms of mean differences
3 generated from studies directly comparing different interventions, together with the results of
4 the NMA (this is from a random effects model, which was chosen on the basis of model fit),
5 in terms of mean differences comparing each available treatment option with placebo/statin.

6 Table 9: Risk differences for percentage change in non-HDL cholesterol at 3-12 7 months; direct pairwise meta-analysis results and NMA results

			NMA random effects model – mean difference (95% credible intervals)	
Intervention	Comparison	Pairwise estimates - mean difference (95% confidence intervals)	Base case model	Model with meta- regression on baseline lipid levels
Standard care	Placebo/ statin	NA	-10.83 (-26.95, 5.80)	-8.84 (-29.11, 11.76)
Ezetimibe		-15.5 (-26.61, -4.39) ^a	-20.02 (-33.05, -6.88)	-3.28 (-79.70, 77.44)
Inclisiran		NA	-45.14 (-58.62, -30.95)	-30.72 (-103.20, 46.14)

			NMA random effects m difference (95% credib	odel – mean le intervals)	
Intervention	Comparison	Pairwise estimates - mean difference (95% confidence intervals)	Base case model	Model with meta- regression on baseline lipid levels	
Evolocumab plus ezetimibe		NA	NA	NA	
PCSK9i		-42.47 (-48.45, -36.50) ^b	-46.99 (-54.26, -39.38)	-30.00 (-104.40, 48.65)	

(a) Fixed effects model 1

2 (b) Random effects model

3 3.4.3 Inconsistency and goodness of fit

4 Both fixed effects and random effects baseline models were fitted to the data from the

5 studies. Large heterogeneity was observed for this outcome, with a large reduction in

6 residual deviance and DIC seen in the RE NMA model, relative to the FE NMA model. This,

7 together with the moderate between-study SD supports choice of the RE model structure.

8 An inconsistency model was run and the model fit statistics were as seen in Table 10. The

9 global check for inconsistency indicates that there is little inconsistency in this dataset, with

10 similar model fit in the RE NMA and RE UME models, and no evidence of local

11 inconsistency.

12 Table 10: Model fit statistics – percentage change in non-HDL cholesterol at 3-12 13 months

Relative effect models ^(a)	Between study SD (95% Crl)	Meta-regression covariate on baseline lipid level	Deviance information criterion (DIC)	Total residual deviance
NMA Fixed effects	-	-	146.1	95.6
NMA (Random effects)	9.44 (5.47, 16.36)	-	74.2	14.3
Unrelated mean- effects (Random effects)	9.92 (5.61, 17.71)	-	74.5	14.4
Meta-regression (Random effects)	11.35 (6.05, 21.43)	-4.14 (-25.56, 16.36)	68.6	12.1

(a) Number of data points in the NMA and inconsistency models (n=14). Number of data points in the metaregression model models (n=12).

14 Figure 4 presents a dev-dev plot, which shows the contributions of each study datapoint to

15 the residual deviance under the random effects UME and NMA models. There is no evidence

16 of inconsistency, as there are no points notably below the line of equality, which would be

17 indicative of data better predicted by the inconsistency model.

18 Taken together, the model fit and dev-dev plots suggest there was little evidence of 19 inconsistency in these data.

20 Figure 6: Contribution of each study datapoint to the residual deviance under the

21 random effects UME and NMA models - percentage change in non-HDL cholesterol at

22 3-12 months



2 3.4.4 Sensitivity analyses and subgroup analyses

3 Within the sensitivity analysis, including studies reporting baseline non-HDL, adding a meta-

4 regression term on baseline non-HDL levels resulted in a model with good statistical fit, but

5 the effect of baseline LDL was again uncertain, -4.14 (95% Crl: -25.56, 16.36), with the 6 interval including zero.

7 3.5 Absolute change from baseline in non-HDL cholesterol

8 3.5.1 Network and data

9 Eight studies^{1, 3, 11, 16, 18-21} were identified as reporting outcome data for absolute change from
10 baseline in non-HDL cholesterol. Five treatments were included in the network: placebo
11 (including placebo and/or statin treatment), standard care, ezetimibe, alirocumab, and
12 evolocumab 420mg. Alirocumab and evolocumab 420 mg were modelled as a single fixed
13 class. The network can be seen in Figure 3 and the trial data for each of the studies included
14 in the NMA are presented in Table 11.

1 Figure 7: Network diagram for absolute change in non-HDL cholesterol



3

4 Table 11: Study data for absolute change in non-HDL cholesterol (mmol/litre) at 3-12 5 months network meta-analysis

			Absolute change LDL-C (mmol/L)	
Study	Intervention	Comparison	Mean differe nce	SE
Cannon, 2015 ³ IMPROVE-IT	Ezetimibe	Placebo	-0.52	0.043
Masuda 2015 ¹⁶	Ezetimibe	Placebo	-0.61	0.263
Ran, 2017 ²¹ NSTE-ACS	Ezetimibe	Placebo	-1.06	0.189
Kereiakes, 2015 ¹¹ ODYSSEY COMBO I	Alirocumab	Placebo	-1.08	0.144
Raber, 2022 ²⁰ PACMAN-AMI	Alirocumab	Placebo	-1.58	0.119
Nicholls, 2016 ¹⁹ GLAGOV	Evolocumab 420 mg	Placebo	-1.64	0.070
Nicholls, 2022 ¹⁸ HUYGENS	Evolocumab 420 mg	Placebo	-1.70	0.189
Ako, 2019 ¹ ODYSSEY J-IVUS	Alirocumab	Standard care	-1.26	0.073

6 3.5.2 Results of network meta-analysis

7 Table 12 summarises the results of the pairwise meta-analyses in terms of mean differences 8 generated from studies directly comparing different interventions, together with the results of

9 the NMA (this is from a random effects model, which was chosen on the basis of model fit),

10 in terms of mean differences comparing each available treatment option with placebo/statin.

1 Table 12: Risk differences for absolute change in non-HDL cholesterol at 3-12 months; 2 direct pairwise meta-analysis results and NMA results

			NMA random effects n difference (95% credib	nodel – mean ble intervals)	
Intervention	Comparison	Pairwise estimates - mean difference (95% confidence intervals)	Base case model	Model with meta- regression on baseline lipid levels	
Standard care	Placebo/ statin	NA	-0.24 (-1.17, 0.70)	-0.17 (-1.33, 1.01)	
Ezetimibe		-0.67 (-1.00, -0.33) ^a	-0.71 (-1.23, -0.21)	0.11 (-3.16, 3.42)	
Inclisiran		NA	NA	NA	
Evolocumab plus ezetimibe		NA	NA	NA	
PCSK9i		-1.45 (-1.67, -1.22) ^a	-1.50 (-1.93, -1.08)	-0.71 (-3.78, 2.41)	
(a) Random effe	cts model				

3 (a) Random effects model

4 3.5.3 Inconsistency and goodness of fit

- 5 Both fixed effects and random effects baseline models were fitted to the data from the
- 6 studies. Moderate heterogeneity was observed for this outcome, with a substantial reduction
- 7 in residual deviance and DIC seen in the RE NMA model, relative to the FE NMA model.
- 8 This, together with the moderate between-study SD supports choice of the RE model
- 9 structure.
- 10 An inconsistency model was run and the model fit statistics were as seen in Table 13. The
- 11 global check for inconsistency, modelling treatment effects independently of each other,
- 12 indicates that there is little inconsistency in this dataset, with similar residual deviance and
- 13 DIC in the RE NMA and RE UME models, and no evidence of local inconsistency.

14 Table 13: Model fit statistics – absolute change in non-HDL cholesterol at 3-12 months

Relative effect models ^(a)	Between study SD (95% Crl)	Meta-regression covariate on baseline lipid level	Deviance information criterion (DIC)	Total residual deviance
NMA Fixed effects	-	-	7.3	24.0
NMA (Random effects)	0.35 (0.11, 0.89)	-	-4.2	8.2
Unrelated mean- effects (Random effects)	0.36 (0.11, 0.90)	-	-4.2	8.2
Meta-regression (Random effects)	0.42 (0.12, 1.22)	-0.22 (-1.10, 0.63)	-4.2	8.0

(a) Number of data points in all models (n=8).

- 15 Figure 8 presents a dev-dev plot, which shows the contributions of each study datapoint to
- 16 the residual deviance under the random effects UME and NMA models. There is no evidence
- 17 of inconsistency, as there are no points notably below the line of equality, which would be
- 18 indicative of data better predicted by the inconsistency model.

19 Taken together, the model fit and dev-dev plots suggest there was little evidence of 20 inconsistency in these data.

- 21 Figure 8: Contribution of each study datapoint to the residual deviance under the
- 22 random effects UME and NMA models absolute change in non-HDL cholesterol at 3-23 12 months



2 3.5.4 Sensitivity analyses and subgroup analyses

Within the sensitivity analysis, including studies reporting baseline non-HDL, adding a metaregression term on baseline non-HDL levels resulted in a small reduction in total residual
deviance, indicating improved model fit, but no change in DIC. The effect of baseline LDL
was again both small and uncertain, -0.22 (95% Crl: -1.1, 0.63), with the interval including
zero.

4 Risk of bias and indirectness

2 An overall risk of bias assessment was conducted for the studies and outcomes included in 3 the NMA. Overall risk of bias for each study-outcome was determined by consideration of the 4 independent domains of bias: selection bias, performance and detection bias, attrition bias, 5 and outcome reporting bias. For each study, if there was no risk of bias in any domain, the 6 risk of bias was given a rating of 'low risk of bias'. If there was risk of bias in just 1 domain, 7 the risk of bias rated as 'some concerns', but if there was risk of bias in 2 or more domains 8 the risk of bias was given a 'high risk of bias' rating.

9 As seen in Table 14, the majority of the relevant evidence for the NMAs had a low risk of
10 bias. For studies where there were some concerns or high risk of bias, this was most
11 frequently due to concerns about selection bias, for example, imbalance in baseline
12 characteristics between groups, or insufficient information about randomisation procedures.
13 Full risk of bias details can be found in the evidence tables and GRADE tables for the

14 pairwise meta-analysis in evidence review D.

Study	% change LDL-C	Absolute change LDL-C	% change non-HDL-C	Absolute change non- HDL-C
Ako, 2019 ¹ ODYSSEY J-IVUS	Some concerns	Some concerns	Some concerns	Some concerns
Arimura, 2012 ²	-	Some concerns	-	-
Cannon, 2015 ⁴ ODYSSEY COMBO II	Some concerns	Some concerns	-	Some concerns
Cannon, 2015 ³ IMPROVE-IT	Low	Some concerns	Low	-
Gao, 2021 ⁵	-	Some concerns	-	-
Giugliano, 2012 ⁶ LAPLACE-TIMI 57a	Low	Low	Low	-
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Low	Low	Low	-
Han, 2020 ⁷ ODYSSEY EAST	Low	- Low		-
Hao, 2022 ⁸	-	High	-	-
Hougaard, 2017 ⁹	Low	Low -		-
Joshi, 2017 ¹⁰	Some concerns	Some - concerns		-
Kereiakes, 2015 ¹¹ ODYSSEY COMBO I	Low	Low	Low	Low
Koh, 2018 ¹² ODYSSEY KT	Low	Low	Low	-
Kouvelos, 2013 ¹³	-	Low	-	-
Luo 2014 ¹⁴	-	Some concerns	-	-
Luo 2016 ¹⁵	-	Some concerns	-	-
Masuda 2015 ¹⁶	High	High	High	High

15 Table 14: Pairwise meta-analysis risk of bias (RoB) assessment per NMA outcome

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Study	% change LDL-C	Absolute change LDL-C	% change non-HDL-C	Absolute change non- HDL-C
McCullough, 2018 ¹⁷ ODYSSEY-LONG TERM	Some concerns	-	Some concerns	-
Nicholls, 2016 ¹⁹ GLAGOV	-	Some concerns	-	Some concerns
Nicholls, 2022 ¹⁸ HUYGENS	-	Low	-	Low
Raber, 2022 ²⁰ PACMAN-AMI	-	Low	-	Low
Ran, 2017 ²¹ NSTE-ACS	Some concerns	Some concerns	-	Some concerns
Ray, 2019 ²² DM-DYSLIPEDIMIA	Some concerns	-	Some concerns	-
Ray, 2019 ²² DM-INSULIN	Low	-	Low	-
Ray, 2020 ²³ ORION-10	Low	Low	Some concerns	-
Ray, 2020 ²³ ORION-11	Low	Low Some concerns		-
Rehberger, 2022 ²⁴	-	High	-	-
Ren, 2017 ²⁵	-	Some -		-
Sabatine, 2017 ²⁶ FOURIER	Low	Low	Low	-
Schwartz, 2018 ²⁷ ODYSSEY OUTCOMES	-	Some concerns	-	-
Tsujita, 2015 ²⁸ PRECISE-IVUS	Low	Low	-	-
Ueda, 2017 ²⁹ ZIPANGU	-	Some concerns	-	-
Wang 2016 ³¹	-	High	-	-
Wang 2017 ³⁰	-	High	-	-
West 2011/2011a ^{32, 33}	-	Low	-	-

1 Key: colour shading represents level of risk (green = low; orange = some concerns; red = high).

2 An assessment of the directness of the evidence was also conducted for the studies and

3 outcomes included in the NMA. Overall directness for each study-outcome was determined

4 by consideration of how well the population, intervention, comparison, and outcomes

5 matched the review protocol.

6 As seen in Table 15, the majority of the relevant evidence for the NMAs had no indirectness.7 For one study there was serious indirectness due to significant imbalance in the statin dose

8 between the treatment groups, while some studies had minor indirectness due to using a

9 weighted mean over a treatment period greater than the protocol specified 12 months. In

10 other studies, insufficient details were provided to assess directness. Full details can be

11 found in the evidence tables and GRADE tables for the pairwise meta-analysis.

1 Table 15: Pairwise meta-analysis indirectness assessment per NMA outcome

Study	% change LDL-C	Absolute change LDL-C	% change non-HDL-C	Absolute change non- HDL-C
Ako, 2019 ¹	No	No	No	No
ODYSSEY J-IVUS	indirectness	indirectness	indirectness	indirectness
Arimura, 2012 ²	-	No indirectness	-	-
Cannon, 2015⁴ ODYSSEY COMBO II	Serious	Serious	-	Serious
Cannon, 2015 ³ IMPROVE-IT	No indirectness	-	No indirectness	-
Gao, 2021 ⁵		No indirectness	-	-
Giugliano, 2012 ⁶ LAPLACE-TIMI 57a	Unclear	Unclear	-	Unclear
Giugliano, 2012 ⁶ LAPLACE-TIMI 57	Unclear	Unclear	-	Unclear
Han, 2020 ⁷ ODYSSEY EAST	-	No indirectness	-	No indirectness
Hao, 2022 ⁸	-	No indirectness	-	-
Hougaard, 2017 ⁹	No indirectness	No indirectness	-	-
Joshi, 2017 ¹⁰	No indirectness	No indirectness	-	-
Kereiakes, 2015 ¹¹ ODYSSEY COMBO I	No indirectness	No indirectness	No indirectness	No indirectness
Koh, 2018 ¹² ODYSSEY KT	No indirectness	No indirectness	No indirectness	-
Kouvelos, 2013 ¹³	-	No indirectness	-	-
Luo 2014 ¹⁴	-	No indirectness	-	-
Luo 2016 ¹⁵	-	No indirectness	-	-
Masuda 2015 ¹⁶	No indirectness	No indirectness	No indirectness	No indirectness
McCullough, 2018 ¹⁷ ODYSSEY-LONG TERM	No indirectness	-	No indirectness	-
Nicholls, 2016 ¹⁹ GLAGOV	-	Minor indirectness	-	Minor indirectness
Nicholls, 2022 ¹⁸ HUYGENS	-	Unclear	-	Unclear
Raber, 2022 ²⁰ PACMAN-AMI	-	No indirectness	-	No indirectness
Ran, 2017 ²¹ NSTE-ACS	No indirectness	No indirectness	-	No indirectness
Ray, 2019 ²² DM-DYSLIPEDIMIA	No indirectness	-	No indirectness	-

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Study	% change LDL-C	Absolute change LDL-C	% change non-HDL-C	Absolute change non- HDL-C
Ray, 2019 ²² DM-INSULIN	No indirectness	-	No indirectness	-
Ray, 2020 ²³ ORION-10	Minor indirectness	Minor indirectness	Minor indirectness	-
Ray, 2020 ²³ ORION-11	Minor indirectness	Minor indirectness	Minor indirectness	-
Rehberger, 2022 ²⁴		Unclear		
Ren, 2017 ²⁵	-	No indirectness	-	-
Sabatine, 2017 ²⁶ FOURIER	No indirectness	No indirectness	No indirectness	-
Schwartz, 2018 ²⁷ ODYSSEY OUTCOMES	No indirectness	No indirectness	-	-
Tsujita, 2015 ²⁸ PRECISE-IVUS	No indirectness	No indirectness	-	-
Ueda, 2017 ²⁹ ZIPANGU	-	Unclear	-	-
Wang 2016 ³¹	-	No indirectness	-	-
Wang 2017 ³⁰	-	No indirectness	-	-
West 2011/2011a ^{32, 33}	-	No indirectness	-	-

1 Key: colour shading represents level of indirectness (green = none; yellow = minor indirectness; orange = serious 2 indirectness; grey = unclear/could not be assessed).

3

4

5 Evidence summary

2 The order of efficacy for reducing cholesterol levels showed PCSK9i to achieve the largest

3 reduction, with inclisiran achieving similar but slightly lower reductions, and ezetimibe

4 achieving clinical important reductions that were considerably lower than that of the

5 injectable therapies. This was true for all analysed outcomes, except percentage change in

6 non-HDL because inclisiran was not present in this network.

7 The results for all outcomes were heterogeneous when using fixed-effects models, so the
8 results were analysed using random effects models, which showed no inconsistency. The
9 majority of the evidence was directly applicable to the review protocol.

10 In the percentage change in LDL-C and absolute change in non-HDL-C networks most of the

11 evidence was at low risk of bias. However, in the absolute change in LDL-C and percentage

12 change in non-HDL-C networks, the majority of the evidence was rated as having some

13 concerns or high risk of bias.

14 For absolute and percentage change in LDL-C, there was little uncertainty for most of the

15 estimates for active treatments compared to placebo, except for inclisiran in the percentage

16 change network and evolocumab plus ezetimibe in the absolute change network. For

17 absolute and percentage change in non-HDL-C there was uncertainty in the network for all 18 interventions.

19

5.1 Modified GRADE assessments

	ble To. Modified GNADE table for NMA data on change from baseline in cholesteror								
No. of studie	f Study s design	Sample size	Effect estimates	Risk of bias	Indirectness	Inconsistency	Quality ¹		
Percenta	Percentage change from baseline in LDL cholesterol								
18	RCT	49,429	See Table 3	No serious	No serious	No serious	High		
Absolute change from baseline in LDL cholesterol									
30	RCT	68,262	See Table 6Table 3	Serious ²	No serious	No serious	Moderate		
Percenta	Percentage change from baseline in non-HDL cholesterol								
13	RCT	35,190	See Table 9	No serious	No serious	No serious	High		
Absolute	Absolute change from baseline in non-HDL cholesterol								
8	RCT	14,744	See Table 12	Serious ²	No serious	No serious	Moderate		

 Table 16: Modified GRADE table for NMA data on change from baseline in cholesterol

¹ Imprecision was not included in ths GRADE assessment but was considered during committee discussions of the evidence, taking into account 95% confidence intervals around the point estimate of the effect, any relevant MIDs, committee expertise and the effect of a single intervention based on multiple outcomes. ²>50% of studies or participants in the NMA judged to have some concerns or high risk of bias. Quality downgraded 1 level. 1

6 Discussion

2 In the networks, the placebo group was acknowledged to involve statin use, and to include
3 studies using statin as the comparator without a placebo control. Consideration was
4 therefore given to pooling the placebo and standard care groups, as was done for the
5 pairwise analyses. However, although we appear to be estimating an extra parameter in
6 'standard care', because the networks connect and the model fit is reasonable, this is
7 justified statistically. These models can be viewed as estimating the effect of the treatment
8 relative to a suitable reference, either placebo or standard care. Additionally, only two
9 treatments, alirocumab and evolocumab, were compared with standard care in trials, in all
10 datasets there was also trial evidence on their effect compared with placebo and
11 inconsistency models indicated no inconsistency.

economic modelling, these models support the use of the percentage change outcome.
Although the absolute data allows more studies and one additional treatment to be included,
the conclusions on treatments' relative effectiveness are unchanged. However, regarding
heterogeneity, the between-study SD was 3.91 SD units (7.287/1.866), and 5.11 SD units
(0.1892/0.03699) for the percentage and absolute models, respectively. This means that the
percentage data were estimated to be more homogeneous than the absolute data and so are
preferred.

Regarding the meta-regression analyses, the large credible intervals around the metaregression term for baseline severity, and the fact that they include zero, mean that it is not possible to determine if there was an effect of baseline lipid level for these data. Given the size of the interval, what is estimated to be a negative effect of baseline severity for non-HDL could feasibly be a positive effect, so mean coefficients from these models could lead to unstable inferences. Furthermore, the projections extend beyond the data modelled – the highest baseline lipid levels reported and modelled were 4.2 mmol/litre, whilst the projection goes up to 6 mmol/litre.

For full discussion and conclusions see the committee's discussion of the evidence in the
evidence review. For linked economic modelling see the separate economic analysis report.

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Appendices

2 Appendix A: WinBUGS Code

3 All codes are derived from Dias, S., Welton, N.J., Sutton, A.J. & Ades, A.E. NICE DSU

4 Technical Support Document 2: A Generalised Linear Modelling Framework for Pairwise and

5 Network Meta-Analysis of Randomised Controlled Trials. 2011; last updated September

6 2016 (available from http://www.nicedsu.org.uk). This work should be cited whenever the

7 code is used whether in its standard form or adapted.

A.18 NMA of contrast between arms, normal likelihood, FE on 9 study, FE for treatment class

```
10 # Normal likelihood, identity link
```

```
11 # Trial-level data given as treatment differences
```

```
12 # Fixed effects model for multi-arm trials
```

13

```
14 ## Updated with a class effect on d
```

```
15 model{ # *** PROGRAM STARTS
```

```
16 for(i in 1:ns2) { # LOOP THROUGH 2-ARM STUDIES
```

- 17 y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
- 18 #Deviance contribution for trial i

```
19 resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
```

```
20
```

}

```
21
```

```
22 for(i in (ns2+1):(ns2+ns3)) { # LOOP THROUGH THREE-ARM STUDIES
```

```
23 for (k in 1:(na[i]-1)) { # set variance-covariance matrix
```

```
24 for (j in 1:(na[i]-1)) {
```

}

}

```
25 Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
```

```
26
```

```
27
```

```
28 Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
```

```
29 # multivariate normal likelihood for 3-arm trials
```

```
30 y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
```

```
31 #Deviance contribution for trial i
```

32 for (k in 1:(na[i]-1)){ # multiply vector & matrix

33 ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]

```
1
        z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
 2
                   }
 3
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
 4
    }
 5
                                  # LOOP THROUGH ALL STUDIES
 6 for(i in 1:(ns2+ns3)){
 7
      for (k in 2:na[i]) {
                              # LOOP THROUGH ARMS
 8
        var[i,k] <- pow(se[i,k],2)</pre>
 9
        prec[i,k] <- 1/var[i,k]
                               # set precisions
10
        delta[i,k] <- d[t[i,k]] - d[t[i,1]]
11
       }
12
   }
13 totresdev <- sum(resdev[])
                                     #Total Residual Deviance
14 d[1]<-0
            # treatment effect is zero for reference treatment
15
16 ## FIXED CLASS MODEL
17 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
18 for (k in nt-nclass+1:nt){ d[k] <- classD }
19 classD ~ dnorm(0, 0.0001)
20
21 ## Dummy vars
22 dv1 <- base m[1,1]
23 dv2 <- base sd[1,1]
24 }
                           # *** PROGRAM ENDS
25
```

A.26 NMA of contrast between arms, normal likelihood, RE on 27 study, FE for treatment class

- 28 # Normal likelihood, identity link
- 29 # Trial-level data given as treatment differences
- 30 # Random effects model for multi-arm trials
- 31 ## Updated with a class effect on d
- 32 model{ # *** PROGRAM STARTS
- 33 for(i in 1:ns2) { # LOOP THROUGH 2-ARM STUDIES

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```
1
      y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
 2 #Deviance contribution for trial i
 3
      resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
 4
     }
                                      # LOOP THROUGH THREE-ARM STUDIES
 5 for(i in (ns2+1):(ns2+ns3)) {
 6
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
 7
         for (j in 1:(na[i]-1)) {
 8
            Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
 9
          }
10
       }
11
      Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
12 # multivariate normal likelihood for 3-arm trials
13
      y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
14 #Deviance contribution for trial i
15
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
16
         ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
17
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
18
                    }
19
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
20
     }
21 for(i in 1:(ns2+ns3)){
                                        # LOOP THROUGH ALL STUDIES
22
      w[i,1] <- 0 # adjustment for multi-arm trials is zero for control arm
23
      delta[i,1] <- 0
                             # treatment effect is zero for control arm
24
      for (k in 2:na[i]) {
                                # LOOP THROUGH ARMS
25
         var[i,k] <- pow(se[i,k],2) # calculate variances</pre>
26
         prec[i,k] <- 1/var[i,k]
                                 # set precisions
27
       }
                                # LOOP THROUGH ARMS
28
      for (k in 2:na[i]) {
29 # trial-specific LOR distributions
30
         delta[i,k] ~ dnorm(md[i,k],taud[i,k])
31 # mean of random effects distributions, with multi-arm trial correction
32
         md[i,k] <- d[t[i,k]] - d[t[i,1]] + sw[i,k]
33 # precision of random effects distributions (with multi-arm trial correction)
```

```
1
        taud[i,k] <- tau *2*(k-1)/k
 2 # adjustment, multi-arm RCTs
 3
        w[i,k] <- (delta[i,k] - d[t[i,k]] + d[t[i,1]])
 4 # cumulative adjustment for multi-arm trials
 5
        sw[i,k] <- sum(w[i,1:k-1])/(k-1)
 6
       }
 7
   }
 8 totresdev <- sum(resdev[])
                                     #Total Residual Deviance
 9 d[1]<-0
               # treatment effect is zero for reference treatment
10
11 ## FIXED CLASS MODEL
12 ## (assuming that trts 2 to (nt-nclass) are not in a class
13 ## and trts nclass to nt are the same class)
14 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
15 for (k in nt-nclass+1:nt){ d[k] <- classD }
16 classD ~ dnorm(0, 0.0001)
17
18 sd ~ dunif(0,upperSD) # vague prior for between-trial SD
19 tau <- pow(sd,-2) # between-trial precision = (1/between-trial variance)
20
21 dv1 <- base m[1,1]
22 dv2 <- base sd[1,1]
23
24 }
                           # *** PROGRAM ENDS
25
```

A.36 NMA of contrast between arms, normal likelihood, FE on 27 study, RE for treatment class

```
28 ## Updated with a class effect on d
29
30 # Normal likelihood, identity link
31 # Trial-level data given as treatment differences
32 # Fixed effects model for multi-arm trials
```

33 model{ # *** PROGRAM STARTS

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```
# LOOP THROUGH 2-ARM STUDIES
 1 for(i in 1:ns2) {
 2
      y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
 3 #Deviance contribution for trial i
 4
      resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
 5
    }
 6
                                     # LOOP THROUGH THREE-ARM STUDIES
 7 for(i in (ns2+1):(ns2+ns3)) {
 8
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
 9
         for (j in 1:(na[i]-1)) {
10
            Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
          }
11
12
       }
13
      Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
14 # multivariate normal likelihood for 3-arm trials
15
      y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
16 #Deviance contribution for trial i
17
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
18
         ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
19
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
20
                    }
21
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
22
     }
23
24 for(i in 1:(ns2+ns3)){
                                    # LOOP THROUGH ALL STUDIES
                                # LOOP THROUGH ARMS
25
       for (k in 2:na[i]) {
26
         var[i,k] <- pow(se[i,k],2)
27
         prec[i,k] <- 1/var[i,k]
                                # set precisions
28
         delta[i,k] <- d[t[i,k]] - d[t[i,1]]
29
       }
30
    }
31 totresdev <- sum(resdev[])</pre>
                                       #Total Residual Deviance
32 d[1]<-0
               # treatment effect is zero for reference treatment
33
```

- 1 ## RE on CLASS MODEL
- 2 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
- 3 for (k in nt-nclass+1:nt){ d[k] ~ dnorm(classD, tauD) }

```
4 classD ~ dnorm(0, 0.0001)
```

```
5 tauD <- pow(sdD,-2)
```

```
6 sdD ~ dunif(0,upperSD) # Vary limits if necessary
```

```
7
```

```
8 ## Dummy vars
```

```
9 dv1 <- base_m[1,1]
```

```
10 dv2 <- base_sd[1,1]
```

```
11 } # *** PROGRAM ENDS
```

```
12
```

A.43 NMA of contrast between arms, normal likelihood, RE on 14 study, RE for treatment class

```
15 # Normal likelihood, identity link
16 # Trial-level data given as treatment differences
17 # Random effects model for multi-arm trials
18 ## Updated with a class effect on d (with RE)
                              # *** PROGRAM STARTS
19 model{
                               # LOOP THROUGH 2-ARM STUDIES
20 for(i in 1:ns2) {
21
      y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
22 #Deviance contribution for trial i
23
      resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
24
    }
25 for(i in (ns2+1):(ns2+ns3)) {
                                    # LOOP THROUGH THREE-ARM STUDIES
26
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
27
         for (j in 1:(na[i]-1)) {
28
           Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
29
          }
30
       }
31
      Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
32 # multivariate normal likelihood for 3-arm trials
33
      y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
```

```
1 #Deviance contribution for trial i
 2
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
 3
         ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
 4
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
 5
                    }
 6
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
 7
    }
                                       # LOOP THROUGH ALL STUDIES
 8 for(i in 1:(ns2+ns3)){
 9
      w[i,1] <- 0 # adjustment for multi-arm trials is zero for control arm
10
      delta[i,1] <- 0
                            # treatment effect is zero for control arm
                                # LOOP THROUGH ARMS
11
      for (k in 2:na[i]) {
12
         var[i,k] <- pow(se[i,k],2) # calculate variances</pre>
13
         prec[i,k] <- 1/var[i,k] # set precisions
14
       }
15
                                # LOOP THROUGH ARMS
      for (k in 2:na[i]) {
16 # trial-specific LOR distributions
17
         delta[i,k] ~ dnorm(md[i,k],taud[i,k])
18 # mean of random effects distributions, with multi-arm trial correction
19
         md[i,k] <- d[t[i,k]] - d[t[i,1]] + sw[i,k]
20 # precision of random effects distributions (with multi-arm trial correction)
21
         taud[i,k] <- tau *2*(k-1)/k
22 # adjustment, multi-arm RCTs
23
         w[i,k] \le (delta[i,k] - d[t[i,k]] + d[t[i,1]])
24 # cumulative adjustment for multi-arm trials
25
         sw[i,k] <- sum(w[i,1:k-1])/(k-1)
26
       }
27
     }
28 totresdev <- sum(resdev[])
                                       #Total Residual Deviance
29 d[1]<-0
               # treatment effect is zero for reference treatment
30
31 ## RE on CLASS MODEL
32 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
33 for (k in nt-nclass+1:nt){ d[k] ~ dnorm(classD, tauD) }
```

15 ## Updated with a class effect on d

```
1 classD ~ dnorm(0, 0.0001)
2 tauD <- pow(sdD,-2)
3 sdD ~ dunif(0,upperSD) # Vary limits if necessary
4
5 sd ~ dunif(0,upperSD) # vague prior for between-trial SD
6 tau <- pow(sd,-2) # between-trial precision = (1/between-trial variance)
7 dv1 <- base_m[1,1]
8 dv2 <- base_sd[1,1]
9
10 } # **** PROGRAM ENDS
11</pre>
```

A.52 NMA of contrast between arms, normal likelihood, FE on 13 study, FE treatment class with additional parameter for 14 high-dose evolocumab (420mg)

```
16
17 # Normal likelihood, identity link
18 # Trial-level data given as treatment differences
19 # Fixed effects model for multi-arm trials
                              # *** PROGRAM STARTS
20 model{
                              # LOOP THROUGH 2-ARM STUDIES
21 for(i in 1:ns2) {
22
      y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
23 #Deviance contribution for trial i
24
      resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
25
    }
26
27 for(i in (ns2+1):(ns2+ns3)) { # LOOP THROUGH THREE-ARM STUDIES
28
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
29
         for (j in 1:(na[i]-1)) {
           Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
30
31
          }
32
       }
33
      Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
```

```
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```

```
1 # multivariate normal likelihood for 3-arm trials
 2
      y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
 3 #Deviance contribution for trial i
 4
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
 5
         ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
 6
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
 7
                    }
 8
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
 9
    }
10
                                   # LOOP THROUGH ALL STUDIES
11 for(i in 1:(ns2+ns3)){
12
                                # LOOP THROUGH ARMS
       for (k in 2:na[i]) {
13
         var[i,k] <- pow(se[i,k],2)</pre>
14
         prec[i,k] <- 1/var[i,k]
                                 # set precisions
15
         delta[i,k] <- d[t[i,k]] - d[t[i,1]]
16
       }
17
     }
18 totresdev <- sum(resdev[])
                                      #Total Residual Deviance
19 d[1]<-0
               # treatment effect is zero for reference treatment
20
21 ## FIXED CLASS MODEL
22 ## (assuming that trts 2 to (nt-nclass) are not in a class
23 ## and trts nt-nclass to nt-1 are a class,
24 ## with treat nt is high dose Evo)
25 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
26 for (k in nt-nclass+1:nt-1){ d[k] <- classD }
27 d[nt]<- classD + evohigh
28 classD ~ dnorm(0, 0.0001)
29 evohigh ~ dnorm(0, 0.0001)
30
31 ## Dummy vars
32 dv1 <- base_m[1,1]
33 dv2 <- base_sd[1,1]
```

1 } # *** PROGRAM ENDS
2

A.63 NMA of contrast between arms, normal likelihood, RE on 4 study, FE treatment class with additional parameter for 5 high-dose evolocumab (420mg)

- 6 ## Updated with a class effect on d
- 7
- 8 # Normal likelihood, identity link
- 9 # Trial-level data given as treatment differences
- 10 # Random effects model for multi-arm trials
- 11 model{ # *** PROGRAM STARTS
- 12 for(i in 1:ns2) { # LOOP THROUGH 2-ARM STUDIES

```
13 y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
```

14 #Deviance contribution for trial i

```
15 resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
```

```
16 }
```

```
17 for(i in (ns2+1):(ns2+ns3)) { # LOOP THROUGH THREE-ARM STUDIES
```

```
18 for (k in 1:(na[i]-1)) { # set variance-covariance matrix
```

```
19 for (j in 1:(na[i]-1)) {
```

}

```
20 Sigma[i,j,k] <- V[i]^*(1-equals(j,k)) + var[i,k+1]^*equals(j,k)
```

21

```
22 }
```

```
23 Omega[i,1:(na[i]-1),1:(na[i]-1)] <- inverse(Sigma[i,,]) #Precision matrix
```

```
24 # multivariate normal likelihood for 3-arm trials
```

```
25 y[i,2:na[i]] ~ dmnorm(delta[i,2:na[i]],Omega[i,1:(na[i]-1),1:(na[i]-1)])
```

```
26 #Deviance contribution for trial i
```

```
27 for (k in 1:(na[i]-1)){ # multiply vector & matrix
```

```
28 ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
```

}

```
29 z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
```

```
30
```

```
31 resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
```

```
32 }
```

```
1
      w[i,1] <- 0 # adjustment for multi-arm trials is zero for control arm
                            # treatment effect is zero for control arm
 2
      delta[i,1] <- 0
                               # LOOP THROUGH ARMS
 3
      for (k in 2:na[i]) {
 4
         var[i,k] <- pow(se[i,k],2) # calculate variances</pre>
 5
         prec[i,k] <- 1/var[i,k] # set precisions
 6
       }
 7
      for (k in 2:na[i]) {
                               # LOOP THROUGH ARMS
 8 # trial-specific LOR distributions
 9
         delta[i,k] \sim dnorm(md[i,k],taud[i,k])
10 # mean of random effects distributions, with multi-arm trial correction
11
         md[i,k] <- d[t[i,k]] - d[t[i,1]] + sw[i,k]
12 # precision of random effects distributions (with multi-arm trial correction)
13
         taud[i,k] <- tau *2*(k-1)/k
14 # adjustment, multi-arm RCTs
15
         w[i,k] <- (delta[i,k] - d[t[i,k]] + d[t[i,1]])
16 # cumulative adjustment for multi-arm trials
17
         sw[i,k] <- sum(w[i,1:k-1])/(k-1)
18
       }
19
    }
20 totresdev <- sum(resdev[])
                                      #Total Residual Deviance
21 d[1]<-0
               # treatment effect is zero for reference treatment
22
23 ## FIXED CLASS MODEL
24 ## (assuming that trts 2 to (nt-nclass) are not in a class
25 ## and trts nt-nclass to nt-1 are a class,
26 ## with treat nt is high dose Evo)
27 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
28 for (k in nt-nclass+1:nt-1){ d[k] <- classD }
29 d[nt]<- classD + evohigh
30 classD ~ dnorm(0, 0.0001)
31 evohigh ~ dnorm(0, 0.0001)
32
33 sd ~ dunif(0,upperSD)
                             # vague prior for between-trial SD
```

10

```
1 tau <- pow(sd,-2) # between-trial precision = (1/between-trial variance)
2 dv1 <- base_m[1,1]
3 dv2 <- base_sd[1,1]
4
5 } # *** PROGRAM ENDS
6</pre>
```

A.77 NMA of contrast between arms, normal likelihood, RE on 8 study, FE for treatment class with meta-regression on 9 baseline lipid level

```
11 # Normal likelihood, identity link
12 # Trial-level data given as treatment differences
13 # Random effects model for multi-arm trials
                              # *** PROGRAM STARTS
14 model{
15
16 for(i in 1:ns2) {
                               # LOOP THROUGH 2-ARM STUDIES
      y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
17
18 #Deviance contribution for trial i
      resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]
19
20
21
     }
22 for(i in (ns2+1):(ns2+ns3)) { # LOOP THROUGH THREE-ARM STUDIES
23
           #V[i] <- pow(se[i,1],2)
24
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
25
         for (j in 1:(na[i]-1)) {
26
           Sigma[i,j,k] <- V[i]*(1-equals(j,k)) + var[i,k+1]*equals(j,k)
          }
27
28
       }
29
      Omega[i,1:2,1:2] <- inverse(Sigma[i,,]) #Precision matrix
30 # multivariate normal likelihood for 3-arm trials
31
      y[i,2:3] ~ dmnorm(delta[i,2:3],Omega[i,1:2,1:2])
32 #Deviance contribution for trial i
33
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
```

```
1
         ydiff[i,k]<- y[i,(k+1)] - delta[i,(k+1)]
 2
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
 3
                     }
 4
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
 5
     }
 6
                                        # LOOP THROUGH ALL STUDIES
 7 for(i in 1:(ns2+ns3)){
 8
      w[i,1] <- 0 # adjustment for multi-arm trials is zero for control arm
 9
      delta[i,1] <- 0
                             # treatment effect is zero for control arm
10
           #V[i] <- pow(se[i,1], 2)
11 #pooled baseline lipid
12 base[i]<- (base m[i,1]+base m[i,2])/2
13
                              # LOOP THROUGH ARMS
14 for (k in 2:na[i]) {
15
         var[i,k] <- pow(se[i,k],2) # calculate variances WAS se[i,k]</pre>
16
         prec[i,k] <- 1/var[i,k]
                                 # set precisions
17
        }
18
      for (k in 2:na[i]) {
                                # LOOP THROUGH ARMS
19 # trial-specific LOR distributions
20
         delta[i,k] ~ dnorm(md[i,k],taud[i,k])
21 # mean of random effects distributions, with multi-arm trial correction
22
         md[i,k] <- d[t[i,k]] - d[t[i,1]] + (beta[t[i,k]] - beta[t[i,1]])*base[i] + sw[i,k]
23 # precision of random effects distributions (with multi-arm trial correction)
24
         taud[i,k] <- tau *2*(k-1)/k
25 # adjustment, multi-arm RCTs
26
         w[i,k] <- (delta[i,k] - (d[t[i,k]] - d[t[i,1]] + (beta[t[i,k]] - beta[t[i,1]])*base[i]))
27 # cumulative adjustment for multi-arm trials
28
         sw[i,k] <- sum(w[i,1:k-1])/(k-1)
29
       }
30
31
     }
32 totresdev <- sum(resdev[])</pre>
                                        #Total Residual Deviance
33
```

```
1 ## Trt eff with class effect
```

```
# treatment effect is zero for reference treatment
 2 d[1]<-0
 3 sd ~ dunif(0,upperSD) # vague prior for between-trial SD
 4 tau <- pow(sd,-2) # between-trial precision = (1/between-trial variance)</p>
 5
 6 ## FIXED CLASS MODEL
 7 ## (assuming that trts 1-3 are not in a class and trts 4-7 are the same class)
 8 for (k in 2:nt-nclass){ d[k] ~ dnorm(0,.0001) }
 9 for (k in nt-nclass+1:nt){ d[k] <- classD }</pre>
10 classD ~ dnorm(0, 0.0001)
11
12 ###Covariate Coefficients. All active equal.
13 beta[1]<-0
14 beta[2]<-0
15 for (k in 3:nt){ beta[k] <- B }
16 B ~ dnorm(0, 0.0001)
17
18 dv1 <- base sd[1,1]
19 }
                           # *** PROGRAM ENDS
20
```

A.&1 Unrelated mean-effects (UME or inconsistency) model of 22 contrast between arms, normal likelihood, RE on study

```
23
```

```
24 ## UME
```

- 26 # Normal likelihood, identity link
- 27 # Trial-level data given as treatment differences
- 28 # Random effects model for multi-arm trials
- 29 model{ # *** PROGRAM STARTS
- 30 for(i in 1:ns2) { # LOOP THROUGH 2-ARM STUDIES
- 31 y[i,2] ~ dnorm(delta[i,2],prec[i,2]) # normal likelihood for 2-arm trials
- 32 #Deviance contribution for trial i
- 33 resdev[i] <- (y[i,2]-delta[i,2])*(y[i,2]-delta[i,2])*prec[i,2]

```
1 }
 2 for(i in 1:(ns2+ns3)){
                                       # LOOP THROUGH ALL STUDIES
 3
      w[i,1] <- 0 # adjustment for multi-arm trials is zero for control arm
 4
      delta[i,1] <- 0
                            # treatment effect is zero for control arm
 5
                               # LOOP THROUGH ARMS
      for (k in 2:na[i]) {
 6
         var[i,k] <- pow(se[i,k],2) # calculate variances</pre>
 7
         prec[i,k] <- 1/var[i,k] # set precisions
 8
       }
 9
10
   for (k in 2:na[i]) {
                             # LOOP THROUGH ARMS
11 # trial-specific mean difference random effects distribution
12
         delta[i,k] ~ dnorm(d[tc[i,1],tc[i,k]],tau)
13
       }
14
    }
15 for(i in (ns2+1):(ns2+ns3)) { # LOOP THROUGH THREE-ARM STUDIES
16
      for (k in 1:(na[i]-1)) { # set variance-covariance matrix
17
         for (j in 1:(na[i]-1)) {
18
           Sigma[i,j,k] <- V[i]^*(1-equals(j,k)) + var[i,k+1]^*equals(j,k)
19
          }
20
       }
21
      Omega[i,1:2,1:2] <- inverse(Sigma[i,,]) #Precision matrix
22 # multivariate normal likelihood for 3-arm trials
23
      y[i,2:3] ~ dmnorm(delta[i,2:3],Omega[i,1:2,1:2])
24 #Deviance contribution for trial i
25
      for (k in 1:(na[i]-1)){ # multiply vector & matrix
26
         ydiff[i,k] <- y[i,(k+1)] - delta[i,(k+1)]
27
         z[i,k]<- inprod2(Omega[i,k,1:(na[i]-1)], ydiff[i,1:(na[i]-1)])
28
                    }
29
      resdev[i]<- inprod2(ydiff[i,1:(na[i]-1)], z[i,1:(na[i]-1)])
30
    }
31
32
33
```

```
1 totresdev <- sum(resdev[]) #Total Residual Deviance
 2
 3 for(i in 1:(ns2+ns3)){
 4
                  for (k in 1:na[i]) { tc[i,k] <- class[t[i,k]]
                                                                       }
 5
                  }
 6
 7 # treatment effect is zero for control arm
 8 for (c in 1:nt) {
                          d[c,c] <- 0 }
 9 # vague priors for treatment effects
10 for (c in 1:(nt-1)) { # priors for all mean treatment effects
11
      for (k in (c+1):nt) {
12
                          d[c,k] \sim dnorm(0,.00001)
13
                          d[k,c] <- -d[c,k]
14
                          }
15
     }
16
17 sd ~ dunif(0,upperSD) # vague prior for between-trial SD
18 tau <- pow(sd,-2) # between-trial precision = (1/between-trial variance)
19 dv1 <- base m[1,1]
20 dv2 <- base sd[1,1]
21 dv3 <- V[1]
22
23 for (n in 1:nt-nclass){
24
           class[n] <- n
25
           }
26 for (n in nt-nclass+1:nt){
27
           class[n] <- nt-nclass+1
28
           }
29
30
31
32 }
                            # *** PROGRAM ENDS
33
```

1